

# EVA: evaluation of protein structure prediction servers

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## ABSTRACT

**EVA (<http://cubic.bioc.columbia.edu/eva/>) is a web server for evaluation of the accuracy of automated protein structure prediction methods. The evaluation is updated automatically each week, to cope with the large number of existing prediction servers and the constant changes in the prediction methods. EVA currently assesses servers for secondary structure prediction, contact prediction, comparative protein structure modelling and threading/fold recognition. Every day, sequences of newly available protein structures in the Protein Data Bank (PDB) are sent to the servers and their predictions are collected. The predictions are then compared to the experimental structures once a week; the results are published on the EVA web pages. Over time, EVA has accumulated prediction results for a large number of proteins, ranging from hundreds to thousands, depending on the prediction method. This large sample assures that methods are compared reliably. As a result, EVA provides useful information to developers as well as users of prediction methods.**

## INTRODUCTION

*Continuous, automated, large data sets, statistical significance.* The goal of EVA is to evaluate the sustained performance of protein structure prediction servers through a

battery of objective measures for prediction accuracy. While the bi-annual CASP (Critical Assessment of Techniques for Protein Structure Prediction) meetings address the question ‘how well can experts predict protein structures with the help of machines?’, the question addressed by EVA is ‘how well can automatic servers predict protein structures?’. Conceptually, this is similar to CAFASP (Critical Assessment of Fully Automated Structure Prediction), but there is a major difference: EVA provides a continuous, fully automatic and statistically more significant analysis of structure prediction servers, whereas CAFASP only covers a limited number of proteins determined in a period of about 4 months in every 2 years: fewer than 10 proteins were available for the non-homology category at CAFASP3 in 2002. This implies that it is—at best—extremely difficult to infer differences of statistical significance from the CAFASP/CASP data sets. For example, the assessor for secondary structure prediction in 2002 concluded that there was no improvement in secondary structure predictions with respect to the CAFASP/CASP in 2000 although the numerical values differed by over six percentage points.

*A tool for developers of prediction methods.* EVA facilitates developers of structure prediction methods to improve their approaches and users of prediction servers to apply methods judiciously. The ranking of each prediction method is analysed and updated on the web every week. Ranking is a non-trivial task because of the non-uniformity in data sets and in the measures for accuracy. Another complication is that methods are compared most reliably when they are tested under identical conditions, i.e. with identical sets of proteins (1–3). Here, we sketch the EVA mechanisms that enable

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**Table 1.** Prediction methods evaluated by EVA

Method	URL	Main developer(s)	References
<b>Comparative modeling</b>			
3D-Jigsaw	<a href="http://www.bmm.icnet.uk/servers/3djigsaw/">http://www.bmm.icnet.uk/servers/3djigsaw/</a>	PA Bates, P Fitzjohn and BC Moreira	(26,27)
CPHModels	<a href="http://www.cbs.dtu.dk/services/CPHmodels/">http://www.cbs.dtu.dk/services/CPHmodels/</a>	S Brunak <i>et al.</i>	(28)
ESyPred3D	<a href="http://www.fundp.ac.be/urbm/bioinfo/esypred/">http://www.fundp.ac.be/urbm/bioinfo/esypred/</a>	C Lambert	(29)
SDSC1	<a href="http://cl.sdsc.edu/hm.html">http://cl.sdsc.edu/hm.html</a>	IN Shindyalov and PE Bourne	(24)
SwissModel	<a href="http://www.expasy.org/swissmod/">http://www.expasy.org/swissmod/</a>	T Schwede, MC Peitsch and N Guex	(30)
<b>Threading/fold recognition</b>			
3D-PSSM	<a href="http://www.sbg.bio.ic.ac.uk/~3dpssm">http://www.sbg.bio.ic.ac.uk/~3dpssm</a>	L Kelley, B Maccallum and M Sternberg	(31)
BLAST	<a href="http://www.ncbi.nlm.nih.gov/BLAST">http://www.ncbi.nlm.nih.gov/BLAST</a>	S Karlin and S Altschul	(32)
FUGUE	<a href="http://www-cryst.bioc.cam.ac.uk/~fugue/">http://www-cryst.bioc.cam.ac.uk/~fugue/</a>	K Mizuguchi	—
Libellula	<a href="http://www.pdg.cnb.uam.es:8081/libellula.html">http://www.pdg.cnb.uam.es:8081/libellula.html</a>	—	—
Prospect	<a href="http://www.aber.ac.uk/~phiwww/prof/">http://www.aber.ac.uk/~phiwww/prof/</a>	Y Xu	(33)
PSI-BLAST	<a href="http://www.ncbi.nlm.nih.gov/BLAST/">http://www.ncbi.nlm.nih.gov/BLAST/</a>	S Altschul <i>et al.</i>	(9)
SAMt99	<a href="http://www.cse.ucsc.edu/research/compbio/HMM-apps/model-library-search.html">http://www.cse.ucsc.edu/research/compbio/HMM-apps/model-library-search.html</a>	K Karplus, C Barrett and R Hughey	(34,35)
Superfamily	<a href="http://supfam.mrc-lmb.cam.ac.uk/SUPERFAMILY/">http://supfam.mrc-lmb.cam.ac.uk/SUPERFAMILY/</a>	J Gough	(36)
<b>Inter-residue contacts</b>			
CORNET	<a href="http://prion.biocomp.unibo.it/cornet.html">http://prion.biocomp.unibo.it/cornet.html</a>	P Fariselli, O Olmea and A Valencia, R Casadio	(10,37,38)
PDGCON	<a href="http://www.pdg.cnb.uam.es:8081/">http://www.pdg.cnb.uam.es:8081/</a>	F Pazos, O Olmea and A Valencia	(13)
CONcons/CONhydro	<a href="http://www.pdg.cnb.uam.es:8081/">http://www.pdg.cnb.uam.es:8081/</a>	F Pazos, O Olmea and A Valencia	(37–39)
<b>Secondary structure</b>			
APSSP2	<a href="http://www.imtech.res.in/raghava/apssp2/">http://www.imtech.res.in/raghava/apssp2/</a>	G Raghava	(40)
Jpred	<a href="http://jura.ebi.ac.uk:8888/">http://jura.ebi.ac.uk:8888/</a>	JA Cuff and GJ Barton	(41)
PHDsec	<a href="http://cubic.bioc.columbia.edu/predictprotein">http://cubic.bioc.columbia.edu/predictprotein</a>	B Rost and C Sander	(42)
PHDpsi	<a href="http://cubic.bioc.columbia.edu/predictprotein">http://cubic.bioc.columbia.edu/predictprotein</a>	D Przybylski and B Rost	(43)
PROF_king	<a href="http://www.aber.ac.uk/~phiwww/prof/">http://www.aber.ac.uk/~phiwww/prof/</a>	M Ouali and R King	(44)
PROFsec	<a href="http://cubic.bioc.columbia.edu/predictprotein">http://cubic.bioc.columbia.edu/predictprotein</a>	B Rost	(45)
PSIpred	<a href="http://insulin.brunel.ac.uk/psiform.html">http://insulin.brunel.ac.uk/psiform.html</a>	D Jones	(46,47)
SAM-T99sec	<a href="http://www.cse.ucsc.edu/research/compbio/HMM-apps/T99-query.html">http://www.cse.ucsc.edu/research/compbio/HMM-apps/T99-query.html</a>	K Karplus, C Barrett and R Hughey	(34,48)
SSpro2	<a href="http://promoter.ics.uci.edu/BRNN-PRED/">http://promoter.ics.uci.edu/BRNN-PRED/</a>	G Pollastri and P Baldi	(49)

(EVA-CM); inter-residue contact prediction (EVA-con); secondary structure prediction (EVA-sec); and threading (EVA-FR). In the following, we sketch the measures for accuracy employed for each category. Note that the detailed definitions of the scores are available through the EVA web sites.

**EVA-CM.** Implements a small number of criteria—arranged hierarchically from coarser to finer—that measure the accuracy of a comparative model. The assessed aspects of a model include fold type, alignment, whole structure, core structure, loops and side-chains. Final ranking is reported using the ‘pairwise’ comparison of prediction servers (3). From May 2000 to January 2003, predictions were collected from five different servers, resulting in 20 957 submitted models for 9050 different PDB chains. On average, 2.3 models were predicted per chain.

**EVA-con.** Evaluates inter-residue contact/distance predictions. A number of servers predict contacts directly, using neural networks of different kinds trained on contact maps (10,11). There are also predictions of contacts based on assembled structures (12). The current evaluation criteria implemented in EVA-con include: (i) accuracy—the number of the correctly predicted contacts divided by the total number of predicted contacts (13); (ii) improvement over random—the calculated accuracy divided by the random accuracy (13); (iii) distance distribution of the predicted contacts—the weighted

harmonic average difference between the predicted contact distance distribution and the all-pairs distance distribution (14); and (iv) delta evaluation—the percentage of correctly predicted contacts that are within a certain number (delta) of residues of the experimental contact, measured along the sequence (15). EVA-con may also be used to evaluate *ab initio*, fold recognition and comparative modelling servers by transforming models into intra-molecular contacts between the corresponding C-beta atoms (C-alpha for Gly) with a 8 Å cut-off.

**EVA-sec.** Evaluates protein secondary structure predictions. Secondary structures are assigned from 3D structures through DSSP (16) and STRIDE (17). EVA-sec measures accuracy by: (i) per-residue accuracy (18) ( $Q_3$ )—percentage of residues correctly predicted in one of the three states (helix, strand or other); (ii) per-segment accuracy (18,19) (*SOV*)—average overlap between segments (methods that get most of the segment cores right are generally more useful than those that get some of the entire segments right); and (iii) accuracy of predicting structural class—percentage of proteins correctly predicted in one of the following classes: all-alpha, all-beta, alpha/beta and others (20,21). Rankings are presented using both the ‘common subset’ and ‘pairwise’ comparison approaches.

**EVA-FR.** Currently evaluates models only for novel sequences (i.e. proteins for which PSI-BLAST searches do not reveal similarity to a known structure). Since there is no

single measure that can comprehensively assess the quality of threading models, EVA-FR employs an array of alignment dependent and alignment independent measures (22–24). For most of the measures used, two aspects of server performance are considered: (i) the ability to produce good models for each target (rank analysis); and (ii) the ability to assign reliable scores to its models, measured through Receiver Operator Characteristics curves (ROC; note this aspect is often referred to with ‘fold recognition’). Methods are ranked through both the ‘common subset’ and ‘pairwise’ comparison approaches.

## DISCUSSION

*EVA provides an automated and continuous evaluation.* Every week, test sequences are automatically submitted to prediction servers and results are evaluated and posted on the EVA web sites. The test sets are constructed so that methods could not have been trained based on the sequences in the test sets. Moreover, the test sets are as large as possible. In addition, the reliability of the comparisons between methods is maximised by using only test sets common to the methods assessed.

*EVA provides supplemental information to CASP.* Since 1994, the development of structure prediction methods has been influenced by the CASP meetings. While EVA uses well-defined numerical criteria to evaluate sustained performance, expert evaluations are still needed to learn what measures are most useful. However, human assessors are not likely to be able to handle many more test sequences than those at CASP. At the same time, there are problems with ranking methods based on test sets that are too small (1–3). EVA rankings are statistically more significant than those at CASP, because EVA assesses prediction methods continuously on as many proteins every month as CASP in 2 years (1). We believe that CASP needs to be supplemented by a large-scale, automated and continuous assessment, such as that by LiveBench (25) (assessment for threading methods only) and EVA. In fact, EVA may replace certain CASP categories in the future. For example, it was proposed at the last 2002 CASP meeting to eliminate secondary structure predictions from CASP. Instead, EVA-sec will replace CASP/CAFASP for users interested in those methods. This decision was partially influenced by the fact that the evaluation of secondary structure prediction methods has matured and this matured analysis has demonstrated beyond doubt that the set of proteins at CASP5 (2002) was not representative and too small.

*EVA allows developers to focus on developing better methods.* The best secondary structure prediction methods have reached a sustained level of 76% accuracy for the last 2 years (2) which indicates a substantial improvement in secondary structure prediction over the last 4 years. While it is always difficult to choose an appropriate set of measures, EVA uses standard criteria that have been largely used by experts in the area. For secondary structure prediction, these criteria are well established. For all other categories, we are currently experimenting with new criteria, others will be incorporated into EVA upon request from users. The precise definitions of the criteria are available on the web. While we can make our

original scripts available upon request, we currently do not have the resources to cast the whole EVA code into a form that guarantees portability or ease-of-use. Overall, EVA allows developers to focus on the development of better methods, rather than on the generally time-consuming evaluation.

*Extension of the EVA framework to other prediction categories.* In principle, the concepts implemented in EVA could and should be generalised to evaluating a larger variety of prediction methods. Often, the problem is the availability of new high-resolution data. We intend to explore extensions that cover the predictions of protein–protein interactions, membrane regions, signal peptides, cleavage sites, structural/functional motifs and sub-cellular localisation.

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