

ModelCIF: An Extension of PDBx/mmCIF Data Representation for Computed Structure Models

Brinda Vallat^{1,2,3*}, Gerardo Tauriello^{4,5}, Stefan Bienert^{4,5}, Juergen Haas^{4,5}, Benjamin M. Webb⁶, Augustin Židek⁷, Wei Zheng⁸, Ezra Peisach^{1,2}, Dennis W. Piehl^{1,2}, Ivan Anischanka⁹, Ian Sillitoe¹⁰, James Tolchard^{11,12}, Mihaly Varadi^{11,12}, David Baker^{9,13}, Christine Orengo¹⁰, Yang Zhang⁸, Jeffrey C. Hoch¹⁴, Genji Kurisu¹⁵, Ardan Patwardhan¹⁶, Sameer Velankar^{11,12}, Stephen K. Burley^{1,2,3,17,18}, Andrej Sali⁶, Torsten Schwede^{4,5}, Helen M. Berman^{1,2,18} and John D. Westbrook^{1,2,3†}

1 - Research Collaboratory for Structural Bioinformatics Protein Data Bank, Rutgers, The State University of New Jersey, Piscataway, NJ 08854, USA

2 - Institute for Quantitative Biomedicine, Rutgers, The State University of New Jersey, Piscataway, NJ 08854, USA

3 - Cancer Institute of New Jersey, Rutgers, The State University of New Jersey, New Brunswick, NJ 08901, USA

4 - Biozentrum, University of Basel, Basel, Switzerland

5 - Computational Structural Biology, SIB Swiss Institute of Bioinformatics, Basel, Switzerland

6 - Department of Bioengineering and Therapeutic Sciences, the Quantitative Biosciences Institute (QBI), and the Department of Pharmaceutical Chemistry, University of California, San Francisco, San Francisco, CA 94157, USA

7 - DeepMind, London, UK

8 - Department of Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, MI 48109, USA

9 - Department of Biochemistry, and Institute for Protein Design, University of Washington, Seattle, WA 98195, USA

10 - Department of Structural and Molecular Biology, UCL, London, UK

11 - AlphaFold Protein Structure Database, European Molecular Biology Laboratory, European Bioinformatics Institute, Hinxton, Cambridge CB10 1SD, UK

12 - Protein Data Bank in Europe, European Molecular Biology Laboratory, European Bioinformatics Institute, Hinxton, Cambridge CB10 1SD, UK

13 - Howard Hughes Medical Institute, University of Washington, Seattle, WA 98195, USA

14 - Biological Magnetic Resonance Data Bank, Department of Molecular Biology and Biophysics, University of Connecticut, Farmington, CT 06030, USA

15 - Protein Data Bank Japan, Institute for Protein Research, Osaka University, Suita, Osaka 565-0871, Japan

16 - Electron Microscopy Data Bank, European Molecular Biology Laboratory, European Bioinformatics Institute, Wellcome Genome Campus, Hinxton, UK

17 - Research Collaboratory for Structural Bioinformatics Protein Data Bank, San Diego Supercomputer Center, University of California, La Jolla, CA 92093, USA

18 - Department of Chemistry and Chemical Biology, Rutgers, The State University of New Jersey, Piscataway, NJ 08854, USA

Correspondence to Brinda Vallat: brinda.vallat@rcsb.org, modelcifwg@wwpdb.org (B. Vallat) [@buildmodels](https://twitter.com/buildmodels) (B. Vallat), [@salilab_ucsf](https://twitter.com/salilab_ucsf) (A. Sali)

<https://doi.org/10.1016/j.jmb.2023.168021>

Edited by Michael Sternberg

Abstract

ModelCIF (github.com/ihmwg/ModelCIF) is a data information framework developed for and by computational structural biologists to enable delivery of *Findable, Accessible, Interoperable, and Reusable (FAIR)* data to users worldwide. ModelCIF describes the specific set of attributes and metadata associated with macromolecular structures modeled by solely computational methods and provides an extensible data

representation for deposition, archiving, and public dissemination of predicted three-dimensional (3D) models of macromolecules. It is an extension of the Protein Data Bank Exchange / macromolecular Crystallographic Information Framework (PDBx/mmCIF), which is the global data standard for representing experimentally-determined 3D structures of macromolecules and associated metadata. The PDBx/mmCIF framework and its extensions (e.g., ModelCIF) are managed by the Worldwide Protein Data Bank partnership (wwPDB, wwpdb.org) in collaboration with relevant community stakeholders such as the wwPDB ModelCIF Working Group (wwpdb.org/task/modelcif). This semantically rich and extensible data framework for representing computed structure models (CSMs) accelerates the pace of scientific discovery. Herein, we describe the architecture, contents, and governance of ModelCIF, and tools and processes for maintaining and extending the data standard. Community tools and software libraries that support ModelCIF are also described.

© 2023 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Brief history of computed structure models (CSMs)

Protein Data Bank (PDB) is the single global repository for three-dimensional (3D) structures of biological macromolecules determined experimentally using macromolecular crystallography (MX), nuclear magnetic resonance (NMR) spectroscopy, and electron microscopy (3DEM). It was established in 1971 as the first open-access digital data resource in biology with seven protein structures.^{1–2} At the time of writing, the archive contained >200,000 structures of proteins, nucleic acids, and their complexes with one another and with small-molecule ligands (e.g., approved drugs, investigational agents, enzyme cofactors). This metric is a testament to the collective efforts and technological advances made by structural biologists working on all inhabited continents. It also highlights a daunting reality—that 99% of protein structure space remains unexplored by experimental methods. Inspired by the work of Anfinsen in 1973,³ computational structural biologists began trying to predict the 3D structure of a protein from its amino acid sequence.

Two distinct approaches for protein structure prediction⁴ have been pursued (Figure 1). The first approach is template-based structure prediction (also known as homology modeling or comparative modeling), in which the structure of an unknown protein (target) is modeled computationally based on the similarity of its amino acid sequence to that of a protein with a known structure (template). Homology modeling is generally successful when template structures from the PDB can be identified and accurately aligned to the target sequence. The second approach is template-free structure prediction, also known as *ab initio* or *de novo* modeling, which can be applied even when reliable structural templates are not available for the protein of interest. In recent years, intramolecular residue-residue con-

tact predictions based on coevolution data⁵ have been successfully applied for template-free structure prediction.⁶

Several automated software tools and web servers support template-based or template-free structure prediction, including, but not limited to, SWISS-MODEL,⁷ Modeller,⁸ ROSETTA,⁹ I-TASSER,¹⁰ QUARK,¹¹ AlphaFold2,¹² and RoseTTAFold.^{13–14} In the Critical Assessment of Techniques for Protein Structure Prediction (CASP14) challenge conducted in 2020,¹⁵ AlphaFold2 demonstrated unprecedented levels of success, an achievement largely enabled by breakthroughs applying machine learning (ML) approaches to protein structure prediction. Following CASP14, another ML-based method, RoseTTAFold, was developed and subsequently applied in combination with AlphaFold2 to predict the structures of hetero-dimeric complexes of eukaryotic proteins.¹⁴ These ML-based structure prediction methods have proven highly successful and are now capable of generating computed structure models (CSMs) with accuracies comparable to that of lower-resolution experimentally-determined structures.¹⁶

Paralleling advances in protein structure prediction methodologies, data resources were established to provide open access to modeled structures. SWISS-MODEL Repository¹⁷ and ModBase¹⁸ house millions of CSMs of proteins generated using SWISS-MODEL or Modeller, respectively. In addition, the ModelArchive, developed at the Swiss Institute of Bioinformatics (SIB, <https://www.modelarchive.org/>), was created to archive and provide stable digital object identifiers (DOIs) for CSMs referenced in publications. ModelArchive includes CSMs which were stored in the PDB before 2006 and has been accepting new depositions since 2013. At the time of writing, the AlphaFold Protein Structure Database (AlphaFold DB)¹⁹ held more than 200 million protein CSMs generated by AlphaFold2. They are freely available and represent virtually all of the protein sequences cataloged in UniProtKB.²⁰

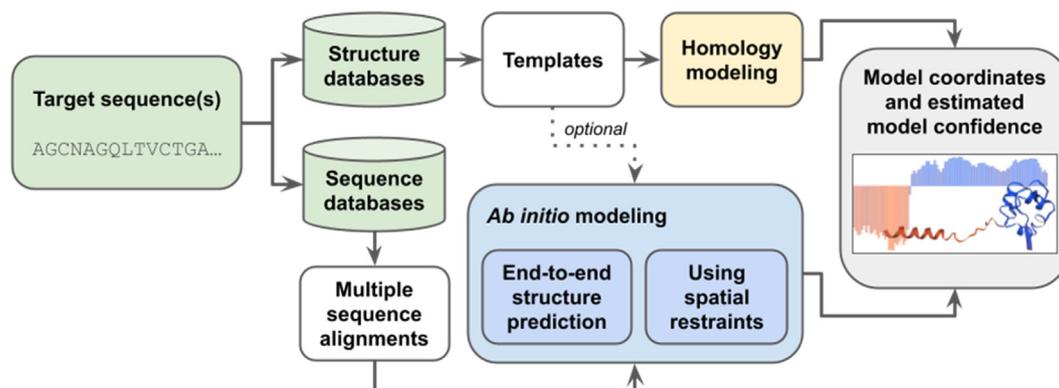


Figure 1. Schematic representation of modeling methods using target sequence(s), structure databases (e.g., PDB), and sequence databases (e.g., Uniclust30⁵¹) as input to produce CSMs and estimates of prediction confidence. Homology modeling uses specific templates as its main input, while *ab initio* methods work without templates. Commonly used *ab initio* methods rely on multiple sequence alignments, which are either used directly as input for end-to-end structure prediction or processed to extract spatial restraints used to generate CSMs.

Significance of data standards in archiving scientific data

Data standards are technical specifications describing the semantics, logical organization, and physical encoding of data and associated metadata. They serve as the foundation for collecting, processing, archiving, and distributing data in a standard format and promoting the *FAIR* (*Findable, Accessible, Interoperable and Reusable*) principles emblematic of responsible data management in the modern era.²¹ In addition to representing the results of a scientific investigation, additional metadata (such as software, authors, citations, references to external data) may be required to support data exchange among different stakeholders, including data generators, archives, and data consumers. If a consistent mechanism is utilized to store such information, it can be shared using common software, agnostic of the data provider, enabling better interoperability among resources and facilitating data search, retrieval, and reuse. Involving community experts in developing and subsequently extending data standards ensures that they are readily adopted by the community and facilitates continuous update of the standards as the field evolves.

History of PDBx/mmCIF data standard for representing macromolecular structures

One of the earliest archival formats in structural biology is the legacy PDB format.²² Developed in the 1970s, it is human and machine readable, easy to parse, and remained the PDB standard exchange format for over forty years. However, it has several drawbacks, including fixed field widths, column positions, and metadata format, which posed severe limitations for archiving large macromolecular structures, data validation, and future expansion to support newer experimental methods.

In 1990, the Crystallographic Information Framework (CIF)²³ was adopted by the International Union of Crystallography (IUCr) as a community data standard to describe small-molecule X-ray diffraction studies. Later, in 1997, the IUCr approved the mmCIF data standard²⁴ to support MX experiments. The original mmCIF data standard was subsequently extended by the PDB to support other experimental methods (e.g., NMR, 3DEM), and to create the PDBx/mmCIF data dictionary.^{25–26} In 2014, this standard was adopted by the worldwide PDB (wwPDB, www.pdb.org)^{2,27} as the master format for the PDB archive. The framework describing PDBx/mmCIF is regulated by Dictionary Definition Language 2 (DDL2), a generic language that supports construction of dictionaries composed of data items grouped into categories.²⁸ DDL2 supports primary data types (e.g., integers, real numbers, text), boundary conditions, controlled vocabularies, and linking of data items together to express relationships (e.g., parent–child relationships). Additionally, software tools have been developed to manage the PDBx/mmCIF dictionary (mmcif.wwpdb.org/docs/software-resources.html). PDBx/mmCIF overcame the limitations of the legacy PDB format and has been extended to represent small-angle solution scattering data²⁹ and integrative structure models.³⁰

History of ModelCIF and the wwPDB ModelCIF Working Group

ModelCIF provides definitions for the specific set of attributes and metadata associated with CSMs. Initial efforts to extend PDBx/mmCIF to support CSMs began in 2001 with creation of the MDB dictionary.³¹ In 2006, the outcomes of a Workshop organized by the Research Collaboratory for Structural Bioinformatics (RCSB) PDB at Rutgers included recommendations to build a common por-

tal for accessing structural models and develop data standards to support CSMs³². The Protein Model Portal (PMP)³³ was created at SIB in collaboration with the Protein Structure Initiative (PSI) Structural Biology Knowledgebase.³⁴ A collaborative project between RCSB PDB and SIB was initiated in 2016 to create data standards that represent CSMs in the PMP and the ModelArchive. These data standards were designed as an extension of PDBx/mmCIF to facilitate interoperability with PDB data. The first set of ModelCIF definitions was released on GitHub in 2018 (github.com/ihmwg/ModelCIF).

The ModelCIF Working Group (WG) was established in 2021 as a collaboration between the wwPDB partners (RCSB PDB, Protein Data Bank in Europe (PDBe), Protein Data Bank Japan (PDBj), Electron Microscopy Data Bank (EMDB), and Biological Magnetic Resonance Bank (BMRB)) and domain experts in computational structural biology ([wwpdb.org/task/modelcif](https://www.pdb.org/task/modelcif)). In addition to wwPDB members, the WG includes representatives from ModelArchive, SWISS-MODEL, Genome3D,³⁵ ModBase, I-TASSER, AlphaFold database, AlphaFold2/DeepMind, and RoseTTAFold. The WG is involved in development and maintenance of the ModelCIF data standard for representing and archiving CSMs and promotes its adoption across the computational biology community. The WG also promotes development of software tools supporting ModelCIF, such as the python-modelcif software library (github.com/ihmwg/python-modelcif). Feedback to the WG via email is welcome (modelcifwg@wwpdb.org).

Results and Discussion

Data definitions reused from PDBx/mmCIF

In developing ModelCIF, various core PDBx/mmCIF dictionary definitions have been reused. These include representation of small-molecule ligands, polymeric macromolecules, biomolecular complexes, and their atomic coordinates, as well as related metadata definitions about modeling software used, bibliographic citations, and author names (Figure 2).

ModelCIF data definitions

Given the variety of existing modeling methods, ModelCIF aims to be flexible regarding data representation. To fulfill this goal, new data categories were introduced to: (i) store input and intermediate results that are of relevance for existing methods; (ii) provide estimates of local and global CSM confidence; (iii) describe steps used to generate CSMs; and (iv) refer to data stored in associated files. New ModelCIF definitions are summarized in Figure 2.

In addition to CSM atomic coordinates, two sets of data items are mandatory: (i) details regarding modeled targets and (ii) list of CSMs included in

the file. New definitions are provided for capturing information pertaining to the origin of modeled molecular entities. This feature is particularly useful for cross-referencing to external databases for macromolecular sequences (e.g., UniProtKB) and small molecules (e.g., PubChem,³⁶ ChEBI).³⁷ Definitions supporting inclusion of small molecules that are not already specified in the wwPDB chemical component dictionary (CCD)³⁸ are also provided.

In ModelCIF, CSMs can be combined into groups that may belong to an ensemble (or cluster). Structural assemblies must be homogeneous (i.e., every CSM in an entry must have identical composition of molecules). Each CSM can be classified as “homology model”, “*ab initio* model” or “other” if neither descriptor is appropriate. The “homology model” category is used for any modeling method (including comparative modeling and protein threading) where the main inputs for generating the CSM are sequence alignments to templates. CSMs generated without templates (or where templates are not considered dominant inputs) are classified as “*ab initio* model” (including fragment sampling and ML-based methods).

Homology modeling methods, as used by SWISS-MODEL and Modeller for example, typically consist of three steps: (i) template identification; (ii) target-template alignment; and (iii) atomic coordinate generation. ModelCIF includes data categories to store the most relevant intermediate results in a standardized way, including a summarized version of the template search results with cross-references to relevant structure databases (e.g., PDB) and detailed information regarding template structures and target-template alignments used for modeling.

Ab initio methods start from sequence information without relying on structural templates. Methods such as I-TASSER generate CSMs using folding simulations guided by deep learning predicted spatial restraints extracted from multiple sequence alignments (MSAs) and corresponding co-evolutionary features. The spatial restraints from deep learning predictors could be residue-residue contacts, distances, dihedral angles, torsion angles, or hydrogen-bonding networks. ModelCIF enables storage of MSAs, homologous templates (optionally used as input structures for *ab initio* methods), and derived spatial restraints, used by *ab initio* folding simulations to model CSMs. ML-based *ab initio* methods such as AlphaFold2 and RoseTTAFold do not rely on features extracted from templates or MSAs but can instead use them as raw input to an “end-to-end” neural network that directly generates the atomic coordinates. Consequently, ModelCIF allows for inclusion of simplified descriptions of relevant input data and intermediate results. ModelCIF can also store information about sequence databases used to construct MSAs (including versions and download

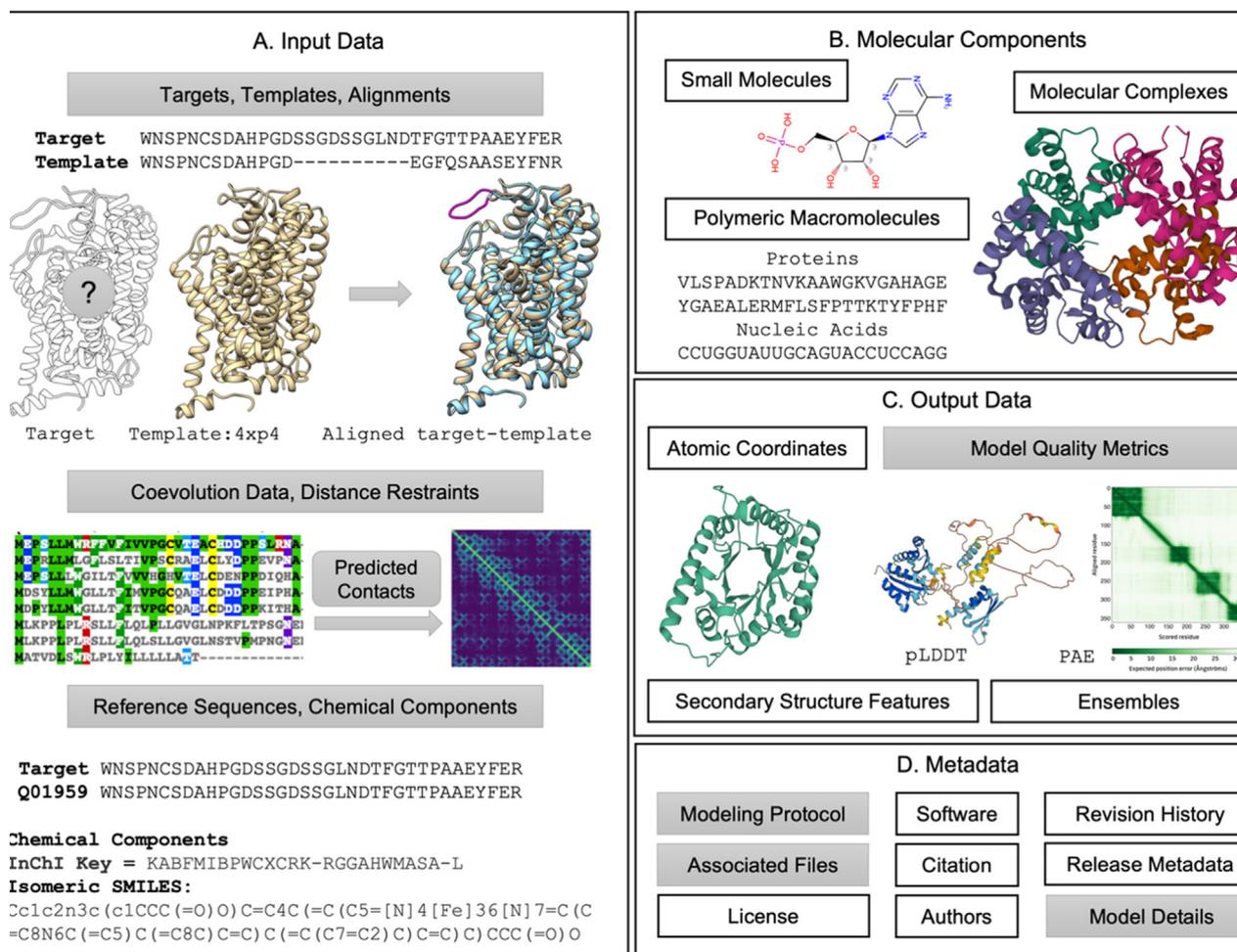


Figure 2. Schematic representation of the data specifications in ModelCIF. Definitions reused from PDBx/mmCIF are shown in white boxes (e.g., Atomic Coordinates) and the newly added definitions are shown in gray boxes (e.g., Model Quality Metrics). (A) Descriptions are provided for input data used in template-based and template-free modeling. (B) Representations of molecular components are retained from PDBx/mmCIF. (C) Definitions for atomic coordinates, secondary structure features, and ensembles are taken from PDBx/mmCIF; descriptions of local and global CSM quality metrics are defined in ModelCIF. (D) Several metadata definitions from PDBx/mmCIF are reused. New metadata definitions regarding modeling protocol, CSM classification (*ab initio*, homology, etc.) and descriptions of associated files are included in ModelCIF. Examples of CSM-specific data and metadata represented in ModelCIF are provided in the [Supplementary Material](#).

URLs) and minimal details regarding any input structures utilized.

While CSMs generated with the newest techniques have become increasingly accurate, it is critical that they are accompanied by estimates of model quality (or prediction confidence). Quality estimates are used to evaluate models and assess their suitability for specific downstream applications. ModelCIF includes flexible support to define any number of quality assessment values. These are classified according to how they are to be interpreted (e.g., probabilities, distances, energies) or as a prediction of the similarity to the correct structure according to well defined metrics such as the TM-score³⁹ or IDDT.⁴⁰ Quality estimate values can be provided globally per CSM and locally

per residue, to identify high- and low-quality regions, and per residue-pair, to enable assessment of contacts and domain orientations.

To facilitate reproducibility of structure prediction and to acknowledge use of publicly available software and web services, ModelCIF allows inclusion of generic definitions describing modeling protocols. Minimally, such definitions may include a free-text description of the modeling protocol as a single step. Ideally, however, multiple steps involved in structure modeling can be described. These steps can be linked to input data (e.g., target sequences, template structures, alignments, predicted contacts), software used (including parameters, version information), and output generated (e.g., CSMs), allowing to capture

intermediate results obtained at each step. To keep data file sizes manageable, ModelCIF provides metadata definitions supporting description of one or more associated files. The data content of associated files can be large intermediate results, such as MSAs or quality estimates for residue-pairs. A variety of generic file formats are allowed for associated files.

Supporting software tools and resources

Table 1 provides a list of software tools and CSM resources that support ModelCIF. Additional details concerning these tools and resources are included in the [Supplementary Material](#).

Advantages of ModelCIF

The value and benefits of ModelCIF are most readily recognized through its support for the *FAIR* principles. ModelCIF provides foundational data standards for archiving CSMs, making them freely available, and enabling seamless data exchange. Moreover, extending PDBx/mmCIF to establish ModelCIF as a data standard in its own right provides the following advantages: (a) existing definitions in PDBx/mmCIF for representing the atomic structures of biological macromolecules, small-molecules, and molecular complexes can be reused; (b) software tools developed to support PDBx/mmCIF can be reused and expanded to support the extension; (c) ModelCIF can be extended to address evolving needs of the

structure prediction community (e.g., protein sequence embedding, neural network model metadata); and (d) the extension facilitates interoperability with other structural biology data resources (e.g., PDB). For example, recent updates to the [RCSB.org](#) web portal to include >1,000,000 CSMs available freely from AlphaFoldDB and the ModelArchive was facilitated by ModelCIF⁴¹. To achieve improved parsing performance and compression, ModelCIF files can be readily converted to BinaryCIF format.⁴²

Conclusion and Perspectives

Computational structural biology is rapidly advancing before our eyes as a discipline. During manuscript preparation, Meta AI announced the development of their own ML-based method for protein structure prediction and used it to generate more than 600 million CSMs that are now publicly available.⁴³ It is also likely that additional open-access resources distributing CSMs of proteins will emerge before this paper appears in print. Ideally, each of these newly established databases of predicted structures will embrace the ModelCIF data standard for deposition, archiving, and dissemination of CSMs. The wwPDB ModelCIF Working Group is committed to maintaining and updating the data standard as new approaches to computational structure modeling of biological macromolecules emerge and are validated. The wwPDB is also supporting community efforts, such as the

Table 1 Software tools and CSM resources supporting ModelCIF.

Software / Resource Name	URL	Description
<i>Software Tools for Reading, Writing, Conversion, and Validation of ModelCIF Files</i>		
python-modelcif	https://github.com/ihmwg/python-modelcif	Software library that supports reading, writing, and validating ModelCIF files and conversion between mmCIF and BinaryCIF
ModelCIF-converters	https://git.scicore.unibas.ch/schwede/modelcif-converters	Collection of ModelCIF conversion tools based on python-modelcif
wwPDB mmCIF software resources webpage	https://mmcif.wwpdb.org/docs/software-resources.html	Website that lists community-developed software libraries and tools that support PDBx/mmCIF, many of which also support ModelCIF (e.g., ciftools-java, py-mmcif)
<i>Modeling Applications and CSM Repositories</i>		
ModelArchive	https://www.modelarchive.org	Repository for CSMs contributed by modelers
SWISS-MODEL ⁷	https://swissmodel.expasy.org	Fully automated protein structure homology modeling server and repository
Modeller ⁸	https://salilab.org/modeller/	Software used for comparative modeling of protein 3D structures
Zhang-Group servers (I-TASSER ¹⁰ , QUARK ¹¹)	https://zhanggroup.org/D-I-TASSER/ https://zhanggroup.org/C-QUARK/	<i>Ab initio</i> and homology modeling servers for protein structure prediction, protein peptide folding, and structure-based function annotation
AlphaFold DB ¹⁹	https://alphafold.ebi.ac.uk	Repository for 3D structures of proteins predicted using AlphaFold2
RoseTTAFold ^{†,13–14}	https://robetta.bakerlab.org	Software tool that uses a three-track neural network to predict protein structures
<i>Visualization Software</i>		
Mol* ⁵²	https://molstar.org	Web-based structure visualization and analysis tool
ChimeraX ⁵³	https://www.cgl.ucsf.edu/chimerax/	Desktop-based structure visualization and analysis tool

† At the time of publication, RoseTTAFold module of the Robetta structure prediction server will support ModelCIF.

3D-Beacons network,⁴⁴ to encourage adoption of common data standards and facilitate access to 3D-structure information.

Looking ahead, CSMs of large, intricately folded ribonucleic acid (RNA) chains may be of particular importance to basic and applied researchers working across fundamental biology, biomedicine, biotechnology/bioengineering, and the energy sciences. Progress in this field is driven by the development of several RNA structure prediction and model quality assessment tools (e.g., SimRNA⁴⁵, RNAComposor⁴⁶, FARFAR2⁴⁷, Vfold⁴⁸, NAST⁴⁹, ARES⁵⁰). Community-organized blind challenges such as CASP will continue to be important in accelerating technical developments in de novo structure prediction for both proteins and nucleic acids.

CRedit authorship contribution statement

Brinda Vallat: Conceptualization, Methodology, Software, Writing – original draft, Visualization, Project administration, Funding acquisition. **Gerardo Tauriello:** Conceptualization, Methodology, Software, Writing – original draft, Visualization. **Stefan Bienert:** Methodology, Software. **Juergen Haas:** Methodology. **Benjamin M. Webb:** Conceptualization, Methodology, Software, Writing – original draft. **Augustin Zidek:** Software, Writing – review & editing. **Wei Zheng:** Software, Writing – review & editing. **Ezra Peisach:** Methodology, Writing – original draft. **Dennis W. Piehl:** Writing – original draft. **Ivan Anischanka:** Software, Writing – review & editing. **Ian Sillitoe:** Writing – review & editing. **James Tolchard:** Writing – review & editing. **Mihaly Varadi:** Writing – review & editing. **David Baker:** Writing – review & editing. **Christine Orengo:** Funding acquisition, Writing – review & editing. **Yang Zhang:** Funding acquisition. **Jeffrey C. Hoch:** Funding acquisition, Writing – review & editing. **Genji Kurisu:** Funding acquisition, Writing – review & editing. **Ardan Patwardhan:** . **Sameer Velankar:** Funding acquisition, Writing – review & editing. **Stephen K. Burley:** Conceptualization, Supervision, Funding acquisition, Writing – original draft. **Andrej Sali:** Funding acquisition, Writing – review & editing. **Torsten Schwede:** Funding acquisition, Writing – review & editing. **Helen M. Berman:** Funding acquisition, Supervision, Writing – review & editing. **John D. Westbrook:** Conceptualization, Methodology, Software, Supervision.

Acknowledgements

The authors thank the tens of thousands of researchers worldwide who enable computational

structure modeling of proteins by depositing ~200,000 experimentally-determined structures to the PDB since 1971. We also gratefully acknowledge contributions to the PDBx/mmCIF data standard made by past members of Worldwide Protein Data Bank partner organizations (RCSB PDB, PDBe, PDBj, EMD, and BMRB) and members of the structural biology community. G.T., S.B., and T.S. acknowledge the contributions of Andrew Mark Waterhouse and Dario Behringer to ModelArchive and support of ModelCIF.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Funding

RCSB PDB core operations are jointly funded by the National Science Foundation (DBI-1832184, PI: S.K. Burley), the US Department of Energy (DE-SC0019749, PI: S.K. Burley), and the National Cancer Institute, the National Institute of Allergy and Infectious Diseases, and the National Institute of General Medical Sciences of the National Institutes of Health (R01GM133198, PI: S.K. Burley). Other funding awards to RCSB PDB by the NSF and to PDBe by the UK Biotechnology and Biological Research Council are jointly supporting development of a Next Generation PDB archive (DBI-2019297, PI: S.K. Burley; BB/V004247/1, PI: Sameer Velankar) and new Mol* features (DBI-2129634, PI: S.K. Burley; BB/W017970/1, PI: Sameer Velankar). B. Vallat acknowledges funding from NSF (NSF DBI-2112966, PI: B. Vallat; NSF DBI-1756248, PI: B. Vallat). A. Sali acknowledges funding from NIH and NSF (NIH R01GM083960, PI: A. Sali; NSF DBI-2112967, PI: A. Sali; NSF DBI-1756250, PI: A. Sali; NIH P41GM109824, PI: M.P. Rout). PDBj is supported by grants from the Database Integration Coordination Program from the department of NBDC program, Japan Science and Technology Agency (JPMJND2205, PI: G. Kurisu), and partially supported by Platform Project for Supporting Drug Discovery and Life Science Research (Basis for Supporting Innovative Drug Discovery and Life Science Research (BINDS)) from AMED under Grant Number 22ama121001. The Protein Data Bank in Europe is supported by European Molecular Biology Laboratory-European Bioinformatics Institute and the AlphaFold Database work is additionally funded by DeepMind. The 3D-Beacons work was supported by funding from the UK Biotechnology and Biological Research Council to PDBe and Christine Orengo group (BB/

S020144/1, BB/S020071/1). J. Hoch acknowledges funding from NIH (R01GM109046) for BMRB. G.T., S.B., and T.S. acknowledge funding from NIH and National Institute of General Medical Sciences (U01GM93324-01), ELIXIR (3D-BiolInfo), and the SIB Swiss Institute of Bioinformatics. Y.Z. acknowledges support from the Extreme Science and Engineering Discovery Environment (XSEDE), which is funded by the National Science Foundation (ACI-1548562). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jmb.2023.168021>.

Received 29 November 2022;
Accepted 16 February 2023;
Available online 23 February 2023

Keywords:

ModelCIF;
PDBx/mmCIF;
Data Standard;
Computed Structure Models;
Protein Structure Prediction

† Deceased.

References

- Protein Data Bank, (1971). Crystallography: Protein Data Bank. *Nature (London) New Biol.* **233**, 223.
- wwPDB consortium, (2019). Protein Data Bank: the single global archive for 3D macromolecular structure data. *Nucleic Acids Res.* **47**, D520–D528.
- Anfinsen, C.R., (1973). Principles that govern the folding of protein chains. *Science* **181**, 223–230.
- Baker, D., Sali, A., (2001). Protein structure prediction and structural genomics. *Science* **294**, 93–96.
- Gobel, U., Sander, C., Schneider, R., Valencia, A., (1994). Correlated mutations and residue contacts in proteins. *Proteins* **18**, 309–317.
- Abriata, L.A., Tamo, G.E., Monastyrskyy, B., Kryshchuk, A., Dal Peraro, M., (2018). Assessment of hard target modeling in CASP12 reveals an emerging role of alignment-based contact prediction methods. *Proteins* **86** (Suppl 1), 97–112.
- Waterhouse, A., Bertoni, M., Bienert, S., Studer, G., Tauriello, G., Gumienny, R., et al., (2018). SWISS-MODEL: homology modelling of protein structures and complexes. *Nucleic Acids Res.* **46** W296 W303.
- Sali, A., Blundell, T.L., (1993). Comparative protein modelling by satisfaction of spatial restraints. *J. Mol. Biol.* **234**, 779–815.
- Leaver-Fay, A., Tyka, M., Lewis, S.M., Lange, O.F., Thompson, J., Jacak, R., et al., (2011). ROSETTA3: an object-oriented software suite for the simulation and design of macromolecules. *Methods Enzymol.* **487**, 545–574.
- Yang, J., Yan, R., Roy, A., Xu, D., Poisson, J., Zhang, Y., (2015). The I-TASSER Suite: protein structure and function prediction. *Nature Methods* **12**, 7–8.
- Mortuza, S.M., Zheng, W., Zhang, C., Li, Y., Pearce, R., Zhang, Y., (2021). Improving fragment-based ab initio protein structure assembly using low-accuracy contact-map predictions. *Nature Commun.* **12**, 5011.
- Jumper, J., Evans, R., Pritzel, A., Green, T., Figurnov, M., Ronneberger, O., et al., (2021). Highly accurate protein structure prediction with AlphaFold. *Nature* **596**, 583–589.
- Baek, M., DiMaio, F., Anishchenko, I., Dauparas, J., Ovchinnikov, S., Lee, G.R., et al., (2021). Accurate prediction of protein structures and interactions using a three-track neural network. *Science* **373**, 871–876.
- Humphreys, I.R., Pei, J., Baek, M., Krishnakumar, A., Anishchenko, I., Ovchinnikov, S., et al., (2021). Computed structures of core eukaryotic protein complexes. *Science* **374** eabm4805.
- Kryshchuk, A., Schwede, T., Topf, M., Fidelis, K., Mout, J., (2021). Critical assessment of methods of protein structure prediction (CASP)-Round XIV. *Proteins* **89**, 1607–1617.
- Shao, C., Bittrich, S., Wang, S., Burley, S.K., (2022). Assessing PDB macromolecular crystal structure confidence at the individual amino acid residue level. *Structure* **30** (1385–1394) e1383.
- Bienert, S., Waterhouse, A., de Beer, T.A., Tauriello, G., Studer, G., Bordoli, L., et al., (2017). The SWISS-MODEL Repository-new features and functionality. *Nucleic Acids Res.* **45** D313 D319.
- Pieper, U., Webb, B.M., Dong, G.Q., Schneidman-Duhovny, D., Fan, H., Kim, S.J., et al., (2014). ModBase, a database of annotated comparative protein structure models and associated resources. *Nucleic Acids Res.* **42** D336 D346.
- Varadi, M., Anyango, S., Deshpande, M., Nair, S., Natassia, C., Yordanova, G., et al., (2022). AlphaFold Protein Structure Database: massively expanding the structural coverage of protein-sequence space with high-accuracy models. *Nucleic Acids Res.* **50** D439 D444.
- UniProt Consortium, (2021). UniProt: the universal protein knowledgebase in 2021. *Nucleic Acids Res.* **49** D480 D489.
- Wilkinson, M.D., Dumontier, M., Aalbersberg, I.J., Appleton, G., Axton, M., Baak, A., et al., (2016). The FAIR Guiding Principles for scientific data management and stewardship. *Sci. Data* **3**, 1–9.
- Westbrook, J.D., Fitzgerald, P.M.D., (2009). Chapter 10 The PDB format, mmCIF formats, and other data formats. In: Bourne, P.E., Gu, J. (Eds.), *Structural Bioinformatics*. Second Edition. John Wiley & Sons Inc, Hoboken, NJ, pp. 271–291.
- Hall, S.R., Allen, F.H., Brown, I.D., (1991). The crystallographic information file (CIF): a new standard archive file for crystallography. *Acta Crystallogr. A* **47**, 655–685.
- Fitzgerald, P.M.D., Westbrook, J.D., Bourne, P.E., McMahon, B., Watenpaugh, K.D., Berman, H.M., (2005). 4.5 Macromolecular dictionary (mmCIF). In: Hall, S.R., McMahon, B. (Eds.), *International Tables for Crystallography G Definition and exchange of*

- crystallographic data. Springer, Dordrecht, The Netherlands, pp. 295–443.
25. Westbrook, J.D., Young, J.Y., Shao, C., Feng, Z., Guranovic, V., Lawson, C., et al., (2022). PDBx/mmCIF Ecosystem: Foundational semantic tools for structural biology. *J. Mol. Biol.* **434**, 167599
 26. Westbrook, J., Henrick, K., Ulrich, E.L., Berman, H.M., (2005). 3.6.2 The Protein Data Bank exchange data dictionary. In: Hall, S.R., McMahon, B. (Eds.), *International Tables for Crystallography*. Springer, Dordrecht, The Netherlands, pp. 195–198.
 27. Berman, H.M., Henrick, K., Nakamura, H., (2003). Announcing the worldwide Protein Data Bank. *Nature Structure Biology*. **10**, 980.
 28. Westbrook, J.D., Berman, H.M., Hall, S.R., (2005). 2.6 Specification of a relational Dictionary Definition Language (DDL2). In: Hall, S.R., McMahon, B. (Eds.), *International Tables for Crystallography*. Springer, Dordrecht, The Netherlands, pp. 61–72.
 29. Malfois, M., Svergun, D.I., (2000). sasCIF: an extension of core Crystallographic Information File for SAS. *J. Appl. Cryst.* **33**, 812–816.
 30. Vallat, B., Webb, B., Westbrook, J.D., Sali, A., Berman, H. M., (2018). Development of a Prototype System for Archiving Integrative/Hybrid Structure Models of Biological Macromolecules. *Structure* **26**, 894–904.
 31. Migliavacca, E., Adzhubei, A.A., Peitsch, M.C., (2001). MDB: a database system utilizing automatic construction of modules and STAR-derived universal language. *Bioinformatics* **17**, 1047–1052.
 32. Berman, H.M., Burley, S.K., Chiu, W., Sali, A., Adzhubei, A., Bourne, P.E., et al., (2006). Outcome of a workshop on archiving structural models of biological macromolecules. *Structure* **14**, 1211–1217.
 33. Haas, J., Roth, S., Arnold, K., Kiefer, F., Schmidt, T., Bordoli, L., et al., (2013). The Protein Model Portal – a comprehensive resource for protein structure and model information. *Database (Oxford)*. bat031.
 34. Gabanyi, M.J., Adams, P.D., Arnold, K., Bordoli, L., Carter, L.G., Flippen-Andersen, J., et al., (2011). The Structural Biology Knowledgebase: a portal to protein structures, sequences, functions, and methods. *J. Struct. Funct. Genomics* **12**, 45–54.
 35. Sillitoe, I., Andreeva, A., Blundell, T.L., Buchan, D.W.A., Finn, R.D., Gough, J., et al., (2020). Genome3D: integrating a collaborative data pipeline to expand the depth and breadth of consensus protein structure annotation. *Nucleic Acids Res.* **48** D314 D319.
 36. Kim, S., Chen, J., Cheng, T., Gindulyte, A., He, J., He, S., et al., (2021). PubChem in 2021: new data content and improved web interfaces. *Nucleic Acids Res.* **49** D1388 D1395.
 37. Hastings, J., Owen, G., Dekker, A., Ennis, M., Kale, N., Muthukrishnan, V., et al., (2016). ChEBI in 2016: Improved services and an expanding collection of metabolites. *Nucleic Acids Res.* **44** D1214 D1219.
 38. Westbrook, J.D., Shao, C., Feng, Z., Zhuravleva, M., Velankar, S., Young, J., (2015). The chemical component dictionary: complete descriptions of constituent molecules in experimentally determined 3D macromolecules in the Protein Data Bank. *Bioinformatics* **31**, 1274–1278.
 39. Zhang, Y., Skolnick, J., (2004). Scoring function for automated assessment of protein structure template quality. *Proteins* **57**, 702–710.
 40. Mariani, V., Biasini, M., Barbato, A., Schwede, T., (2013). IDDT: a local superposition-free score for comparing protein structures and models using distance difference tests. *Bioinformatics* **29**, 2722–2728.
 41. Burley, S.K., Bhikadiya, C., Bi, C., Bittrich, S., Chao, H., Chen, L., et al., (2023). RCSB Protein Data Bank (RCSB.org): Delivery of Experimentally-Determined PDB Structures Alongside One Million Computed Structure Models of Proteins from Artificial Intelligence/Machine Learning. *Nucleic Acids Res.* **51** D488 D508.
 42. Sehnal, D., Bittrich, S., Velankar, S., Koča, J., Svobodová, R., Burley, S.K., et al., (2020). BinaryCIF and CIFTools—Lightweight, Efficient and Extensible Macromolecular Data Management. *PLoS Comput. Biol.* **16** e1008247.
 43. Lin, Z., Akin, H., Rao, R., Hie, B., Zhu, Z., Lu, W., et al., (2022). Evolutionary-scale prediction of atomic level protein structure with a language model. *bioRxiv*. <https://doi.org/10.1101/2022.07.20.500902>.
 44. Varadi, M., Nair, S., Sillitoe, I., Tauriello, G., Anyango, S., Bienert, S., et al., (2022). 3D-Beacons: decreasing the gap between protein sequences and structures through a federated network of protein structure data resources. *GigaScience* **11** giac118.
 45. Boniecki, M.J., Lach, G., Dawson, W.K., Tomala, K., Lukasz, P., Soltysinski, T., et al., (2016). SimRNA: a coarse-grained method for RNA folding simulations and 3D structure prediction. *Nucleic Acids Res.* **44** e63.
 46. Biesiada, M., Purzycka, K.J., Szachniuk, M., Blazewicz, J., Adamiak, R.W., (2016). Automated RNA 3D Structure Prediction with RNAComposer. *Methods Mol. Biol.* **1490**, 199–215.
 47. Watkins, A.M., Rangan, R., Das, R., (2020). FARFAR2: Improved De Novo Rosetta Prediction of Complex Global RNA Folds. *Structure* **28** (963–976) e966.
 48. Li, J., Zhang, S., Zhang, D., Chen, S.J., (2022). Vfold-Pipeline: a web server for RNA 3D structure prediction from sequences. *Bioinformatics* **38**, 4042–4043.
 49. Jonikas, M.A., Radmer, R.J., Laederach, A., Das, R., Pearlman, S., Herschlag, D., et al., (2009). Coarse-grained modeling of large RNA molecules with knowledge-based potentials and structural filters. *RNA* **15**, 189–199.
 50. Townshend, R.J.L., Eismann, S., Watkins, A.M., Rangan, R., Karelina, M., Das, R., et al., (2021). Geometric deep learning of RNA structure. *Science* **373**, 1047–1051.
 51. Mirdita, M., von den Driesch, L., Galiez, C., Martin, M.J., Soding, J., Steinegger, M., (2017). Uniclust databases of clustered and deeply annotated protein sequences and alignments. *Nucleic Acids Res.* **45** D170 D176.
 52. Sehnal, D., Bittrich, S., Deshpande, M., Svobodova, R., Berka, K., Bazgier, V., et al., (2021). Mol* Viewer: modern web app for 3D visualization and analysis of large biomolecular structures. *Nucleic Acids Res.* **49** W431 W437.
 53. Pettersen, E.F., Goddard, T.D., Huang, C.C., Meng, E.C., Couch, G.S., Croll, T.I., et al., (2021). UCSF ChimeraX: Structure visualization for researchers, educators, and developers. *Protein Sci.* **30**, 70–82.